DETERMINATION OF THE threo- AND erythro- CONFIGURATIONS OF VICINAL DIASTEREOMERIC DERIVATIVES OF HIGHER FATTY ACIDS WITH THE AID OF PMR SPECTRA

I. L. Kuranova and L. V. Balykina

UDC 547.396+547.710

PMR spectroscopy is used successfully in the study of the sterochemistry of diastereomeric compounds in which two chiral centers lead to the existence of three and erythre forms. Thus, for example, it has been shown that for symmetrical and unsymmetrical α -glycols [1, 2], β -diols [3], γ -glycols [4], amino alcohols [5, 6], and β -hydroxy esters [7] the chemical bonds of the methine protons of the erythre isomers are located in a weaker field than those of the three isomers, i.e., the condition δ CH(erythre) > δ CH(three) is satisfied. However, this relationship is not always observed. In individual cases — for example, for disubstituted butanes [8] and derivatives of 1,2-diphenylpropane [9] — there is not clear correlation between the configuration and the δ CH value, and the relative magnitude of $\Delta\delta = \delta$ CH(erythre) — δ CH(three) has a positive or negative value according to the nature of the substituent.

Other cases are known in which the chemical shifts of the methine protons of diasteromeric compounds are practically identical, although their spin-spin coupling constants differ, and here again there is no direct correlation between the configuration and the value of J. For such compounds as substituted 1,2-dihalopropanes, 1,2-diphenylpropanes [9], and others [8, 11], Jerythro > Jthreo, and for acyloxylated or alkoxylated 2-amino-1,2-diphenylethanes [12], 2-dimethylamino-1,2-diphenylethanols [13], and other compounds [2, 3], Jerythro < Jthreo.

Information on the PMR spectra of diastereomeric derivatives of higher fatty acids is sparse. The chemical shifts of the protons of individual groups have been given for some diastereomeric pairs: threo- and erythro-6,7-dihydroxy-, -9,10-dibromo- [14], and -9-chloro-10hydroxyoctadecanoic acids [14, 15], and threo-9,10-dihydroxyoctadecanoic acid [14].

In the present paper we give the results of an investigation by the PMR method of position isomers of threo- and erythro-dihydroxy-, acetoxy-hydroxy, and -halohydroxyoctadecanoic and -docosanoic acids and their trimethylsilyl derivatives.



In the PMR spectra of the above-mentioned compounds in the 3-5-ppm region there are two multiplets which are due to the two nonequivalent protons H^{α} (CH $^{\alpha}$ OH) and H^{β} (CH $^{\beta}$ X). The assignment of the signals of the methine protons was made on the basis of a comparison of the PMR spectra of the corresponding dihydroxy-, acetoxyhydroxy-, acetoxybromo-, and halohydro-octadecanoic and -docosanoic acids taken under standard conditions.

Table 1 gives the values of the chemical shifts of the protons in the PMR spectra of the diastereomeric compounds studied. It must be mentioned that the PMR spectra of the individual position isomers and their mixtures are identical, as we showed for the case of the halohydroxy acids as examples.

A. A. Zhdanov Leningrad State University. Translated from Khimiya Prirodnykh Soedinenii, No. 3, pp. 299-305, May-June, 1978. Original article submitted July 11, 1977.

247

TABLE 1. Chemical Shifts in the PMR Spectra of Diastereomeric Derivatives of Higher Fatty Acids (δ , ppm, 100 MHz, benzene)

a b β c d $CH_3(CH_2)_nCH-CH(CH_2)_{m-1}$ CH_2COOR $X=OH, OCOCH_3, Br, C1, at m=4, n=10$ I $R=H, CH_3$ $m=7, n=7$ X OH $m=11, n=7$										
		Chemical shift								
Compound	Config- uration	đ	q	с	CH ^a	$\frac{\Delta\delta CH^{\alpha}}{-\delta CH^{\alpha}} = \frac{-\delta CH^{\alpha}}{erythro}$	CH ³	$2\delta CH^{\beta} =$ = $\delta CH^{\beta} -$ = δCH^{β} ery thro	q	000 C II3
Methyl esters of dihydroxy acids										
Methyl 6,7-dihydroxy- octadecanoate	erythro threo	0,92 0,91	1,35 1,35	$2,21 \\ 2,16 \\ 16$	3,58 3,33	0,25			3,40 3,38	
Methyl 9,10-dihydroxy- octadecanoate	erythro threo	0,93 0,92	$1,33 \\ 1,32$	$2,15 \\ 2,16$	3,55 3,39	0,16			3,39 3,39	
Methyl 13,14-dihy- droxyoctadecanoate	erythro threo	0,92 0,93	1,33 1,33	$2,15 \\ 2,17$	3,57 3,35	0,22			3,38 3,39	
	Acetoxyl) vdro	i xv ae	rids	I I		1 1		1 1	
6(7)-Acetoxy-7(6)-	erythro	0.93	1,32	2,19	3,71	0.17	5,01			1,85
hydroxyoctadecanoic	threo	0,91	1,29	2, 18	3,54	0,17	4,95			1,83
hydroxyoctadecanoic	threo	0,89	1,27	2,11	3,53	0,16	4,98			1,83
13(14)- Acetoxy-14(13)- nydroxydocosanoic acid	erythro threo	0,91 0,91	1,29 1,28	2,15 2,16	3,71 3,60	0,11	5.05 5,01		1	1,81 1,82
	Ha	lohyd	roxy	ació	ls		1 1		1 1	
6(7)-Bromo-7(6)-hy-	erythro	0 94	1,34	2,15	3,56	0.22	3,96	0.13		
droxyoctadecanoic 7-Bromo-6-budroxy-	threo	0,93	1,34 1,34	2,15 2,15	3,34 3,54	0,22	3,83 3,95	0,10		
octadecanoic acid	three	0,93	1,33	2,16	3, 32	0,22	3,83	0,12		
6-Bromo-7-hydroxy-	erythro three	0,93	$1,32 \\ 1,33$	2,14	3,57 3,33	0,24	3,93 3,81	0,12		
9(10)-Bromo-10(9)-hy-	erythro	0,93	1,30	2,16	3,61	0.21	4,00	0.09		
droxyoctadecanoic	threo	$0,94 \\ 0,93$	1,31 1,29	2,17 2,16	$3,40 \\ 3,60$	0,21	3,91 3,99	0,00		
octadecanoic acid	three	0,94	1,31	2,15	3,38	0,22	3,88	0,11		
9-Bromo-10-hydroxy-	erythro threo	0,93 0,94	1,31 1 31	2,14 2,16	3,60	0,23	4,00 3.89	0,11		
13(14)-Bromo-14(13)-	erythro	0,94	1,29	2,17	3,62	0.22	4,02	0.12		
hydroxydocosanoic	threo	0,93	1,29	2,16	3,40	0,22	3,90	-,		
6(7)-Chloro-7(6)-hy-	erythro	0,92	1,31	2.14	3,54	0.12	3,77	0.10		
droxyoctadecanoic	threo erythro	0.93 0.92	1.30 1.31	$2,15 \\ 2.14$	$3,42 \\ 3,52$	0.10	3.77	0.00		
octadecanoic acid	threo	0,93	1,30	2,15	3,42	0,10	3,68	0,09		
6- Chioro-7-hydroxy- octadecanoic acid	erythro threo	0.92 0.91	1.32 1.30	2,10 2,09	3,30 3,43	0,13	3,63	0,09		
9(10)-Chloro-10(9)-hy-	erythro	0,92	1,27	2,14	3,62	0,09	3,84 3,73	0,11		
droxydecanoic acid	ery thro	0,91	1,25	2,13 2,14	3,60	0.11	3,82	0.09		
octadecanoic acid	threo	0,92	1,27	2,16 2 15	3,49 3,61	0,12	3,73 3 81	0,00		
octadecanoic acid	threo	0,92	1.26	2,14	3,52	0,09	3,71	0,10		
13(14)- Chloro- 14(13)- hydroxydocosanoic acid	erythro threo	$0.92 \\ 0.93 \\ 0.93 \\ 0.01 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ 0.02 \\ $	1,28 1,29	2,15 2,15	3,62 3,53	0,09	3,83 3,75	0,08		

It follows from an analysis of the facts presented that for the dihydroxy, halohydroxy, and acetoxyhydroxy acids and their esters the signal of the $C\underline{H}^{\alpha}$ (OH) methine proton for the erythro form is shifted downfield in comparison with the three form, i.e., $\delta(C\underline{H}^{\alpha}OH)$ erythro > $\delta(C\underline{H}^{\alpha}OH)$ three as in the case of α -glycols and β -diels [1-3]. The relative shift $\Delta\delta(CH^{\alpha}OH) = \delta$ erythro — δ three amounts to 8-11 Hz for the chlorohydroxy, 10-17 Hz for the acetoxyhydroxy, 16-20 Hz for the dihydroxy, and 20-22 Hz for the bromohydroxyoctadecanoic and -docosanoic acids (and esters).

The value of $\delta CH(OCOCH_3)$ for all series of acetoxyhydroxy acids is approximately the same, amounting to 5-5.09 ppm. For the series of halohydroxy acids the signals of the δCH^{β} -(Hal) protons of the erythro derivatives are found in lower fields than those of the three

derivatives, and $\Delta\delta(CH^{B}Hal) = \delta_{erythro} - \delta_{threo}$ has approximately the same value for the bromohydroxy and for the chlorohydroxy acids, amounting to 8-11 Hz.

A comparison of the chemical shifts of the $CH^{\beta}(Hal)$ protons shows that the values of $\delta CH^{\beta}(Br)$ (3.87-4.06 ppm) are somewhat greater than $\delta CH^{\beta}(C1)$ (3.72-3.89 ppm). Thus, for the halohydroxy acids no correlation is observed between the electronegativity of the substituent (C1, Br) and the values of $\delta CH(Hal)$, just as for the case of the 2,3-dibromo- and 2,3-dichlorobutanes [8] and the halogen-substituted 1,2-diphenylethanes [16].

The difference in the chemical shifts of the $CH^{\alpha}(OH)$ and $CH^{\beta}(X)$ (X = OH, Cl, Br) methine protons for the erythro and three derivatives is apparently explained by the fact that in the erythro form each of the methine protons experiences the descreening action of β substituents to a greater degree than in the three form.

When an alcoholic hydroxyl in the compounds investigated is replaced by an $OSi(CH_3)_3$ group, the values of $\delta CHOSi(CH_3)_3$ for the three and erythre forms scarcely differ for the case of the acetoxyhydroxy acids, while in the case of the dihydroxy acids for the three form the signal is shifted downfield somewhat, $\Delta\delta CHOSi(CH_3)_3 = \delta_{three} - \delta_{erythree}$ amounting to 5-10 Hz. The greatest difference is observed in the PMR spectra of the silylated diastereomeric bromehydroxy acids: The CHOSi(CH_3)_3 and CHBr methine protons of the three derivatives give individual multiplets, and for the erythree derivative these protons give a common unresolved multiplet shifted downfield. The relative change in the chemical shift $\Delta\delta CHOSi (CH_3)_3 = \delta_{erythre} - \delta_{three}$ amounts to 9-13 Hz. Thus, the PMR spectra of the TMS derivatives permit a clear conclusion to be drawn concerning the configuration only in the case of the bromehydroxy acids, and for the TMS derivatives of the dihydroxy and acetoxyhydroxy acids the differences in the PMR spectra of the three and erythree forms are small (Table 2).

However, when the PMR spectra of the dihydroxy-, acetoxyhydroxy-, and halohydroxyoctadecanoic and -docosanoic acids and their TMS derivatives are compared (see Tables 1 and 2), an interesting relationship is observed. For all the compounds investigated, when a CHOH group

> TABLE 2. Chemical Shifts in the PMR Spectra of the Trimethylsilyl-Substituted Diastereomeric Derivatives of Higher Fatty Acids (δ , ppm, 100 MHz, benzene)

> > $\begin{array}{cccc} \mathbf{c} & \mathbf{d} & \mathbf{g} & \mathbf{f} & \mathbf{d} & \mathbf{c} & \mathbf{b} \\ \mathrm{CH}_3(\mathrm{CH}_3)_n & \mathrm{CH}_{--}\mathrm{CH} (\mathrm{CH}_3)_{m-1} & \mathrm{CH}_2 & \mathrm{COOSi} & (\mathrm{CH}_3)_3; \\ & & & & \\ & & & & \\ & & & & \\ & & & & \mathbf{a} \end{array}$

X=OSi(CH₃)₃, OCOCH₃ Br; at m=4, n=10m=7, n=7

	m	1	í		n =
	m		T	,	11

	Con- Chemical shift								
Derivative	tigura-	a	b	C	d	e	f	g	i
TMS-6,7-dihydroxyocta- decanoic acid TMS-9,10-dihydroxy- octadecanoic acid TMS-13,14-dihydroxy- docosanoic acid TMS-6(7)-acetoxy-7(6)- hydroxyoctadecanoic acid TMS-9(10)-acetoxy-10(9)-	erythro threo erythro threo erythro threo erythro threo erythro	0,18 0,18 0,22 0,18 0,22 0,18 0,16 0,18 0,16	0,26 0,28 0,26 0,26 0,26 0,26 0,27 0,27 0,27	0,91 0,91 0,92 0,91 0,93 0,90 0,92 0,92 0,92	1.31 1,32 1,32 1.31 1.31 1,30 1.31 1,32	2,21 2,23 2,20 2,18 2,20 2,20 2,20 2,18 2,20 2,19	3,60 3,70 3,66 3,72 3,68 3,73 3,73 3,73 3,73 3,78	5,03 5,06 5,07	1,83 1,80
hydroxyoctadecañoic acid	threo	0,20	0,27	0,92	1,31	2,19	3,82	5,11	1,81
14(13)-hydroxydocasano- ic acid	threo	0.21	0,28	0,92 0,92	1,29	2,18	3,85	5,09	1,81
TMS-6(7)-bromo-7(6)- hydroxyoctadecanoic acid	erthro threo	$\begin{array}{c} 0.15 \\ 0.08 \end{array}$	0,25 0,25	0.91 0,91	1,28 1,29	2,15 2,13	3 3,75	84 3,89	
TMS-9(10)-bromo-10(9)- hydroxyoctadecanoic	erthro threo	0,17 0,11	0.24 0,25	0.90 0,91	$\substack{1.27\\1.28}$	$\begin{array}{c} 2.15\\ 2.14 \end{array}$	3,80 3,80	0 3,96	·
TMS-13(14)-bromo- 14(13) hydroxydocosa- noic acid	erthro threo	0,17 0,10	$\substack{0.25\\0.24}$	0,92 0,91	1.27 1 2ð	2.17 2,17	3,9 3,82 j	95 3,98	

TABLE 3. Difference in the Values of the Chemical Shifts of the Methine Protons in the PMR Spectra of the Dihydroxy, Acetoxyhydroxy, and Bromohydroxy Acids and Their TMS derivatives for the three and erythro Forms (δ , ppm, 100 MHz, benzene)

		Chemical shift				
Compound	Configura- tion	снон	CHOSI(CH_),	$\Delta \delta = \delta CHOSI(CH_{3})_{3} - \delta CHOH$		
Methyl 6,7-dihydroxyoctadecano-	erythro	3,58				
até	ervthro	0,00	3.60	0.02		
1 MS=6, 7= dillydroxyoctadecalloic	three		3,70	0,37		
Methyl 9,10-dihydroxyocta-	erythro	3,55				
decanoate	threo	3,39	2.00	0.00		
TMS-9,10-dihydroxyoctadecanoic	erythro		3,00	0,03		
acid	amithro	3 57	0,12	0,00		
ate	three	3,35	No. 1997			
TMS-13.14-dihvdroxydocosanoic	erythro		3,68	0,11		
acid	threo	0.71	3,73	0,38		
6(7)-Acetoxy-7(6)-hydroxy-	erythro	3.71				
octadecanoic acid	threo	0,04	3.77	0.06		
TMS-6(7)-acetoxy-7(6)-hydroxy-	three		3,78	0,24		
9(10)-Acetoxy-10(9)-hydroxy-	erythro	3,69				
decanoic acid	threo	3,53	0.00	0.14		
TMS-9(10)-acetoxy-10(9)-	erythro		3,83	0.14		
hydroxydocosanoic acid	threo	3 71	0,0-	0,20		
13(14)-Acetoxy-14(13)-hydroxy-	three	3.60				
COCOSERCIC a CIU = 14(13) = 13(14) = 2000 = 14(13) = 13(14) = 2000 = 14(13) = 12(14) = 2000 = 14(13) = 12(14) = 2000 = 12(14) = 2000 = 12(14) = 2000 = 2000 = 12(14) = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 2000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 20000 = 200000 = 200000 = 200000 = 200000 = 2000000 = 200000 = 2000000 = 20000000 = 200000000	ervthro	-,	3,86	0,15		
hydroxydocosanoic acid	threo	1.0.00	3,85	0,25		
6(7)-Bromo-7(6)-hydroxyocta-	erythro	3,56				
decanoic acid	threo	1 3,34	3.84	0.28		
TMS 6(7)-bromot(6)-nydroxy-	three		3.75	0,41		
Octadecanoic aciu	erythro	3,60				
octadecanoic acid	threo	3,40		0.20		
TMS- bromo-10(9)-octa-	erythro	j –	3,50	0,30		
decanoic acid	i tirreo	3 69	5,30	0.10		
13(14)-Bromo-14(13)-hydroxy-	three	3.40				
docosanoic acid	ervthro	0,10	3,95	0,32		
I MO-10(14)-DIUMO-14(10)-	threo		3,82	0,42		
nyuroxyuocosanoic aciu		1	.1 ¹	1		

is replaced by a CHOSi(CH₃)₃ group the signal of the methine protons shifts downfield, i.e. δ CHOSi(CH₃)₃ - δ CHOH > 0, and in all cases the greatest value of this change is found for the three derivatives (Table 3).

In the PMR spectra of the TMS derivatives of all the compounds investigated a sharp singlet of the methyl protons of the $COOSi(CH_3)_3$ group was observed at 0.29-0.33 ppm. The position of the second singlet of the methyl protons of the $CHOSi(CH_3)_3$ group for the TMS derivatives of the three and erythro-dihydroxy- and -acetoxyhydroxy acids was approximately the same — at δ 0.23-0.27 ppm — but in the case of the halohydroxy acids for the three derivatives the values of $\delta CHOSi(CH_3)_3$ were shifted somewhat upfield as compared with the erythro derivatives, $\Delta\delta CHOSi(CH_3)_3 = \delta$ erythro — δ three amounting to 14-16 Hz.

An analysis of the PMR spectra of the diastereomeric dihydroxy-, acetoxyhydroxy-, and halohydroxyoctadecanoic and -docosanoic acids and their TMS derivatives shows that the difference in the chemical shifts of the vicinal methine protons may serve as a good criterion for determining the threo and erythro configurations of higher fatty acid derivatives.

EXPERIMENTAL

The PMR spectra were taken on a Varian HA-100 D/15 (100 MHz) instrument in benzene. TMS was used as internal standard. The accuracy of the measurements of the chemical shifts was 0.01-0.02 ppm. The concentration of the solutions was 8-10%. We had synthesized the threoand erythro-halohydroxy acids previously [17], and the threo- and erythro-dihydroxy- and -acetoxyhydroxy acids were obtained in accordance with previous work [18]. The trimethylsilyl derivatives of the corresponding compounds were obtained by treating them with hexamethyldisilazane and chlorotrimethylsilane (2:1) in pyridine solution [19]. The acids were methylated with diazomethane in ethereal solution at room temperature.

The analytical results for all the compounds obtained corresponded to the calculated figures.

CONCLUSIONS

The PMR spectra of vicinal diastereomeric dihydroxy-, acetoxyhydroxy-, and halohydroxyoctadecanoic and -docosanoic acids and their trimethylsilyl derivatives have been considered. It has been shown that the signals of the methine protons of the erythro isomers appear at lower fields than those of the threo isomers. On passing from the dihydroxy, acetoxyhydroxy, and halohydroxy acids to their TMS derivatives, the signal of the methine proton shifts downfield, and $\Delta \delta = \delta CHOSi(CH_3)_3 - \delta CHOH$ is greater for the threo form than for the erythro form. The difference in the chemical shifts of the vicinal methine protons of the dihydroxy, acetoxyhydroxy, and halohydroxy acids and their TMS derivatives can be used to determine the threo and erythro configurations of the corresponding diastereometric derivatives of higher fatty acids.

LITERATURE CITED

- 1. J. Wiemann, G. Dana, Sa-le-Thi-Thuan, and M. Brami, Compt. Rend., 258, 3724 (1964).
- 2. G. Dana, J. Chuche, and M.-R. Monot, Bull. Soc. Chim. Fr., 3308 (1967).
- 3. M. J. P. Maffrand and P. Maroni, Tetrahedron Lett., <u>48</u>, 4201 (1969).
- 4. A. Zysman, G. Dana, and J. Wiemann, Bull. Soc. Chim. Fr., 1019 (1967).
- 5. G. G. Lule and L. K. Keefer, J. Org. Chem., <u>31</u>, 3921 (1966).
- 6. J. W. Hufmann and H. P. Elliott, J. Org. Chem., <u>30</u>, 365 (1965).
- 7. J. Cancell, J. J. Basselier, and J. Jacques, Bull. Soc. Chim. Fr., 1906 (1963).
- 8. A. A. Bothner-By and C. Naar-Colin, J. Am. Chem. Soc., 84, 743 (1962).
- 9. C. A. Kingsbury and D. C. Best, J. Org. Chem., 32, 6 (1967).
- 10. H. Schmid, Can. J. Chem., <u>46</u>, 3415 (1968).
- 11. H. Bodot, J. Fedière, G. Pouzard, and L. Pujol, Bull. Soc. Chim. Fr., 3260 (1968).
- 12. M. E. Munk, M. K. Meilahn, and P. Franklin, J. Org. Chem., 33, 3480 (1968).
- 13. G. Drefahl, G. Heublein, and E. Linke, Z. Chem., 8, 174 (1968).
- 14. R. Kannan, M. R. Subbaram, and K. T. Achaya, Fette, Seifen, Anst., 76, 344 (1974).
- 15. M. Ketola, Acta Chem. Scand., <u>27</u>, 1328 (1973).
- 16. C. A. Kingsbury and W. B. Thornton, J. Am. Chem. Soc., 88, 3159 (1966).
- 17. I. L. Kuranova and L. V. Balykina, Zh. Org. Khim., 9, No. 2, 260 (1973).
- I. L. Kuranova, Yu. D. Shenin, and G. V. Pigulevskii, Zh. Obshch. Khim., <u>31</u>, 3142 (1961);
 <u>32</u>, 1675 (1962); <u>33</u>, 2980 (1963); <u>34</u>, 3487 (1964).
- 19. G. G. Abbot, F. D. Gunstone, and S. D. Hoyes, Chem. Phys. Lipids, 4, 351 (1970).